# A SURVEY OF BONE INTEGRITY FROM CULL SOWS IN IRELAND

W.F. Ryan <sup>1,2</sup> P.B. Lynch <sup>1</sup> J.V. O' Doherty <sup>2</sup>

<sup>1</sup> Department of Pig Production Teagasc, Moorepark Research Centre, Fermoy, Co. Cork, Ireland <sup>2</sup> School of Agriculture, Food Science and Veterinary Medicine. University College Dublin, Dublin 4. Ireland.

# ABSTRACT

The objective of this study was to examine the differences in sow metacarpal properties through various parities and to compare the incidence of locomotory problems between stalled and loose-housed sows. Metacarpals (n=110) of sows from six farms were collected at slaughter and stored at -20°C. Bones from one forelimb of sows in stalls (n = 36) and loose-housed (n= 20) were collected and their articular surfaces examined for Osteochondrosis Dissecans (OCD), these sows were also scored for lameness pre-slaughter. Metacarpals were CT scanned for cross sectional area and moment of inertia. Cylindrical sections from the diaphysis were used for mechanical testing and calculating bone strength indices. The results show there was little change in bone mineral status throughout the range of parities examined and overall metacarpal integrity was not compromised by multiple production cycles. There were no differences detected in the incidence of lameness or joint pathology between housing systems.

# **KEYWORDS**

Pigs, Bone, Parity, Lameness, Osteochondrosis-dissecans

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Changes in modern pig reproduction have led to increased numbers of pigs born throughout a sow's lifetime (Lawlor and Lynch 2007). Such improvements necessarily increase metabolic demands on the reproducing sow. To date there has been no monitoring of how the sows' skeleton has reacted to these changes in Irish conditions. Furthermore, there is no information on how these changes in productivity have affected bone integrity of breeding sows under Irish conditions despite average culling rates of 11% for locomotory problems (Boyle *et al.*, 1998). The purpose of this study was:

- 1. To examine differences in physical characteristics of metacarpals among sows of varying parities from commercial farms; and,
- 2. To compare the articular surfaces in the bones of cull sows in different housing systems and to examine any associations with lameness.

There are numerous reports in the literature citing lameness as a significant cause of premature culling. However, the reason for lameness and any association with housing type remains unclear. High culling rates due to a potentially painful condition may indicate compromised welfare for the animals involved and warrants further detailed examination.

# MATERIALS AND METHODS

The animals used in this survey were those being routinely culled and sent to the abattoir from the herds for commercial reasons. All of the farms commenced routine removal of breeding sows once their eighth parity was reached unless it was removed prematurely for health or economic reasons. No sow was kept beyond their ninth parity. III or severely injured sows were not included in the study as they were not transported to the abattoir but were humanely destroyed on the farm of origin. Four of the farms had an on-farm milling system in place and none of the commercial farms used phytase. Most of the farms fed a wet diet to the breeding sows. Prior to slaughter, sows were observed walking and trotting freely on concrete and lameness scored using the system (0 to 5) of Main et al. (2000). Joint surfaces were grossly examined post mortem for defects consistent with osteochondrosis dissecans (OCD) or osteoarthritis (OA) using the system of Nakano et al. (1984),(1 to 7). Lameness distribution (from loose or stalled systems) percentages were calculated for both the total number of lame and normal sows. Metacarpals were collected from sows from farms A to MP in Table 1 (metacarpals have been used by several authors as indicators of skeletal

#### Table 1 Summary of farm details.

Farm	Number of sows	Housing	Diet type	Diet	Breed <sup>y</sup>	Phytase
A	880	Stalled	Wet	Purchased	LW x LR	None
В	420	Stalled	Wet	Purchased	LW x LR	None
C	550	Stalled	Wet	Home	LW x LR	None
D	2600	Loose	Wet	Home	LW x LR	None
E	2000	Stalled	Wet	Home	LW x LR	None
MP <sup>z</sup>	270	Stalled	Wet	Home	LW x LR	Included
F <sup>x</sup> (Loose)	300	Loose	Dry	Purchased	LW x LR	None

<sup>2</sup>MP= Moorepark Research Farm

<sup>y</sup>LW =Large White LR= Landrace

#### \*Loose housed in groups of 100

#### "Number of sows refers to the number of breeding sows normally kept on farm

health (Crenshaw et *al.* 1981, Combs et *al.*1991, Shaw et *al.* 2006)). The samples were obtained immediately after slaughter and stored at -20°C in polythene bags. Sow carcass weights were also recorded.

One forelimb from 36 Moorepark sows and 20 from a loose-housed farm (F) were also collected. Sows from this farm were Large White \*Landrace crosses and were kept on straw bedding in groups of 100. Sows were typically culled after seventh parity. Joint surfaces of the humerus, radius and ulna were examined on the day of slaughter. Metacarpals were CT scanned in a dorso-palmar position with the bone phantom (Osteo, Siemens, Erlangen, Germany) in the field of view. Scans were taken at the middiaphysis perpendicular to the main axis. CT measurements of the metacarpals were obtained in air as described by Bhatavadekar et al. (2006) with a Somatom Spiral CT scanner (Siemens, Erlangen, Germany) at 130 kV tube voltage, 63mA tube current, 1.9 s scan time and 0° gantry tilt. The field of view was 45cm x 45cm with a pixel matrix of 512 x 512, the slice thickness was 3mm (Figure 1). The



Figure 1. CT image of metacarpal cross section used to measure cross sectional area and moment of inertia.

CT data were processed to produce Jpeg images using VG Studio max1.2 (Volume Graphics GmbH Heidelberg Germany) software and the subsequent images were analysed to measure cross sectional area and moment of inertia using Image J 1.38u (Rasband, W.S., ImageJ, U.S. National Institutes of Health, Bethesda, Maryland, USA). The use of CT scans to measure geometrical properties is preferred to assuming certain shape types and applying standard formulae because assumptions about cross sectional geometry, bone shape and distribution are circumvented (Bhatavadekar *et al.* 2006).

Mid–diaphyseal sections from the metacarpals were cut using a band saw (Rexon Laser, Rexon Ind. Corp. Ltd.) for compression testing. The section had a width to length ratio of 2:1 to reduce the buckling effect as recommended by Keaveny *et al.* (1993).

The principle of Archimedes was used to determine the volume of the bone specimen with marrow and periosteum tissues manually removed. The bone was weighed using an OHAUS Scout pro SPU202 balance, OHAUS Corp. NJ USA in air and suspended in distilled water.

Bone specimens were tested in compression at 5mm min<sup>-1</sup> until failure using a Hounsfield instrument (model H50K-S, Redhill, Surrey, UK). The resulting data were collected using software (QMAT 4.52 S Series) which plotted a graph of applied load (Newtons) versus displacement (mm). Platens were fixed at the lower end only to allow self-alignment. Young's modulus was calculated from the linear section of the stress-strain curve.

Metacarpal ash content was determined using a muffle furnace at 105°C for 12 hours to remove moisture and at 650°C for six hours to determine bone ash percentage on both a dry and fresh weight basis. Bone mineral density was calculated by dividing the ash weight (g) by the specimen volume (cm<sup>3</sup>).

#### **STATISTICS**

Statistical analysis was carried out using the methods of SAS 1985 Inc, Cary, N. Carolina. All data were tested for normality using proc univariate. To analyse the metacarpal data the sows were grouped according to three parity ranges; 1 to 3, 4 to 6 and 7 and over. Differences in

#### Table 2. Summary of average metacarpal values per parity range

PARITY RANGE	Early	Mid	Late	Р	SE <sup>z</sup>			
	(1 - 3)	(4 - 6)	(7 +)					
No. of sows	18	46	46					
Cross sectional area cm <sup>2</sup>	2.12b	2.33a	2.19ba	<0.01	0.023			
Cross sectional moment of inertia mm <sup>4</sup>	1.67b	1.73a	1.75a	<0.001	0.009			
Apparent density g cm <sup>-3</sup>	0.65b	0.90a	0.78a	<0.05	0.019			
Bone mineral density g cm <sup>-3</sup>	0.30	0.30	0.30	NS	0.002			
Elastic Modulus GPa <sup>v</sup>	18.94	18.98	19.68	NS	0.483			
Dry matter %	84.75	85.16	85.31	NS	0.178			
Ash % (fresh basis)	50.13	52.11	52.88	NS	0.416			
Ash % (dry basis)	59.17	61.26	62.01	NS	0.507			

Data presented as least square means, values with subscripts are significantly different

#### <sup>Y</sup>GPa: Giga pascals

metacarpal properties among the three parity ranges were determined using the analysis of variance (ANOVA). Sow carcase weight (Kg) was used as a covariate when analysing parity differences.

Forelimb joint pathology and lameness scores were analysed between housing systems using proc npar1way. A Chi square analysis was performed on the lameness percentages between the two housing systems using proc freq.

## RESULTS

Metacarpal strength and geometrical results for each parity range are presented in **Table 2**. Significant differences among the ranges were found with respect to cross sectional area (P<0.01), cross section moment of inertia (P<0.001) and apparent density (P<0.05). Theses three indices increased significantly from the early to the mid parity range and there were no significant changes thereafter.

There were no significant changes in the other indices measured throughout the parity ranges examined. Specifically, there were no significant decreases in bone mineral density, elastic modulus or ash content from early to late parity in the sows examined.

A seventh farm (F), not included in the metacarpal integrity survey, was used to compare joint pathology between stalled and loose-housed systems where sows are kept on straw bedding. Results from these analyses (**Table 3**) show no significant differences between housing systems for severity of joint surface or lameness scores. A Chi squared analysis showed no significant differences between the housing systems for lameness frequency.

# DISCUSSION

The results (**Table 2**) show geometrical parameters cross section area (CSA) and cross section moment of inertia (CSMI) were significantly different among parity ranges. Parameters related to the quality of the bone substance, namely bone mineral density, elastic modulus, dry matter and ash content, were not significantly different. An initial

increase followed by a plateau in the CSA and CSMI through early to late parity was expected due to the sow growing in size and reaching maturity. In addition to the normal metacarpal growth patterns of size and shape there may also be a different control mechanism for the consistency of bone substance. Size and shape may be secondary properties, which may be altered according to the individual's age and weight-bearing requirements. Tanck et al. (2001) reported a time lag between density and architecture formation in trabecular bone of pigs. In addition, this author stated that architecture was more efficient in later years at adapting to loading and that density is adapted to external load from the early phase of growth. This phenomenon has been found in humans examined by Cheng et al. (2007). This suggests a possible compensation mechanism for lower BMD by increasing bone volume and size. These findings would suggest that larger bones are not necessarily stronger than smaller bones. This study found the metacarpal geometry to reach its maximum dimensions' during the mid parity range, while there was no significant changes in the bone quality parameters. This suggests the bones reached their maximum size and quality during the mid range and there

# Table 3 Median joint<sup> $\gamma$ </sup> and lameness<sup>z</sup> scores [median, (min-max)] for stalled and loose-housed sows

	n=20 Loose	n=36 Stalled	Sig <sup>x</sup>
Humeral head	5.0 (2.0-7.0)	5.0 (1.0-6.0)	NS
Medial condyle	2.0 (1.0-6.0)	2.0 (1.0-7.0)	NS
Lateral condyle	2.0 (1.0-7.0)	2.0 (1.0-5.0)	NS
Aconeal process	1.5 (0.0-6.0)	2.0 (1.0-7.0)	NS
Proximal ulna	2.5 (1.0-5.0)	2.0 (1.0-6.0)	NS
Proximal radius	2.0 (1.0-5.0)	2.0 (1.0-5.0)	NS
Lameness score	1.0 (0-2)	1.0 (0-3.0)	NS

<sup>z</sup>Lameness scoring system 0 = normal gait, 5 = severe lameness

<sup>Y</sup>Joint scores 1 = normal to 7 = severe joint pathology

<sup>x</sup>Sig = Significance: NS no significance (p> 0.05)

<sup>&</sup>lt;sup>z</sup>SE: Standard error

were no significant reductions subsequently in their later parities.

No evidence was found of reducing metacarpal integrity due to multiple pregnancies. Bone mineral density, elastic modulus, dry matter and ash content showed no significant changes from early-to-late parity throughout the sows' lifetime. This demonstrates that sows from a number of different farms are able to maintain metacarpal integrity despite multiple parities and lactations. These conclusions were also made by Giesemann et al.(1998) who found metacarpals and coccygeal vertebrae of mature sows to have higher ash weights than those of young sows. It is widely accepted that a pregnancy and subsequent lactation increases the mineral demands on the sow (Mahan and Fetter 1982). As these minerals are ultimately obtained from the diet it would suggest that sows may adapt to the demands of the foetus and lactation by altering their absorption of dietary minerals. This has also been stated by Nimmo et al. (1981). In addition, during lactation higher feed intakes further increase mineral intake. Furthermore. larger mature sows could consume more feed than younger sows. It is possible that there is a hormonal effect on mineral metabolism and this has been postulated by Kemme et al. (1997) and Vajda et al.(2001) who stated that the female reproductive cycle substantially alters mineral metabolism. It is likely that a combination of these three mechanisms allows the sow to adapt to the higher demands of pregnancy and lactation to maintain longterm bone integrity. Both Phillips et al. (2000) and Vajda et al.(2001) stated that osteoporosis in pregnancy is a rare clinical problem. It is important however to limit the conclusions of the survey to the bones examined as not all bones are effected equally. Mahan and Fetter (1982) found the bones of the axial skeleton to be more susceptible to demineralisation. Maxson and Mahan (1986) stated that this was due to their greater proportion of trabecular bone. Any decreases in bone mass during lactation are transient and appear to be partially restored in humans after weaning. Studies have shown lactation not to increase fracture risk and may even have a protective effect in both rats and humans (Henderson et al. 2000, Vajda et al. 2001). Maxson and Mahan (1986) stated that a relatively short 21-day lactation over two parities in sows did not impose a severe enough nutritional insult to cause differences in bone demineralisation. Vajda et al.(2001) showed reductions in vertebral compressive strength to be restored eight weeks post-lactation in rats and in addition that restoration of BMD in humans is rapid and any decrease in biomechanical parameters associated with lactation rarely cause pathological fractures. Spencer (1979) stated that swine are similar to humans in bone structure and mineral metabolism.

No differences in lameness score or joint pathology were detected between the sows on straw bedding (farm F) and stalled animals (farm MP). All 56 sows examined (**Table 3**) had lesions of OCD and subsequent osteoarthritis. The diagnosis of OCD in such a large proportion has been found by several other authors. Nakano *et al.* (1987) found

greater than 90% and Jørgensen and Vestergaarad (1990) stated OCD was present in all fast growing pigs. OCD is frequently implicated as the main cause of leg weakness and subsequent economic losses from premature culling (Dewey et al. 1993, Kirk et al. 2005, Crenshaw 2006). The chronic persistence of a pathological condition contributing to economic losses would suggest that the elimination of OCD is expensive and potentially difficult. Even today. the exact aetiology remains unclear. Crenshaw (2006) describes OCD as idiopathic. Ekman and Carlson (1998) describe it as multifactorial. In addition to other authors, Schenck et al. (2008), found no difference between confined and stalled sows and Uhlorn et al.(1995) found no difference in the incidence of OCD between purebred pigs and those crossed with wild boar. Both of these studies would indicate that neither genetics nor housing alone were responsible for OCD.

The loose-housed sows had access to more space but that does not imply they were more active as straw bedding would be more comfortable than a concrete floor. In addition, the sows on straw are better insulated from any heat loss to the concrete (Tuyttens, 2005). Heat loss has been shown to affect pig performance and weight gain as this increases metabolic rate and energy requirements. In contrast, exercise causes an increase in muscle mass (Marchant and Broom 1996). This author also demonstrated that increased forelimb muscle weight in the pig has significant influence on bone strength and this was found to occur more in loose-housed sows than with stalled animals. Crenshaw (2006) stated that physical weight bearing influences the incidence of OCD but there was no evidence to support or reject a specific housing type with regard to their influence on OCD. This was also found by Kirk et al.(2005). In contrast, Fredeen and Sather (1978) found cartilage damage to be greater in pigs confined individually compared to those kept in a group. This study found no differences in the incidence of OCD between the housing systems. This would suggest changing pigs from a stalled-house to a loose-housed system alone would not decrease the incidence of OCD.

## CONCLUSIONS

There is no reduction in metacarpal bone integrity with increasing parity. Housing type did not influence joint pathology or incidence of lameness in sows.

- Bhatavadekar, N.B., Daegling, D.J. and Rapoff, A.J.
  2006. Application of an image-based weighted measure of skeletal bending stiffness to great ape mandibles
  American Journal of Physical Anthropology. 131: 243-251.
- Boyle, L., Leonard, F.C., Lynch, B. and Brophy, P. 1998. Sow Culling Patterns and Sow Welfare. Irish Veterinary Journal incorporating Irish Veterinary Times 51: 354-357.
- **Cheng, X., Li, J., Lu, Y., Keyak, J. and Lang, T. 2007.** Proximal femoral density and geometry measurements by quantitative computed tomography: Association with hip fracture. Bone 40: 169-174.

- Combs, N.R., Kornegay, E.T., Lindemann, M.D., Notter, D.R., Wilson, J.H. and Mason, J.P. 1991. Calcium and phosphorus requirement of swine from weaning to market weight: II. Development of response curves for bone criteria and comparison of bending and shear bone testing. J. Anim Sci. 69: 682-693.
- **Crenshaw, T.D. 2006.** Arthritis or OCD Identification and Prevention. Advances in Pork Production 17: 199.
- Crenshaw, T.D., Peo, E.R., Jr., Lewis, A.J. and Moser, B.D. 1981. Bone Strength as a Trait for Assessing Mineralization in Swine: a Critical Review of Techniques Involved. J. Anim Sci. 53: 827-835.
- Dewey, C.E., Friendship, R.M. and Wilson, M.R. 1993. Clinical and postmortem examination of sows culled for lameness. Can Vet J. 34: 555-556.
- **Ekman, S. and Carlson, C. 1998.** The pathophysiology of osteochondrosis. Vet Clin North Am Small Anim Pract 28: 17-32.
- **Fredeen, H. and Sather, A. 1978.** Joint damage in pigs reared under confinement. Canadian Journal of Animal Science 58: 759-773.
- Giesemann, M.A., Lewis, A.J., Miller, P.S. and Akhter, M.P. 1998. Effects of the reproductive cycle and age on calcium and phosphorus metabolism and bone integrity of sows. J. Anim Sci. 76: 796-807.
- Henderson, P.H., Sowers, M.F., Kutzko, K.E. and Jannausch, M., L. 2000. Bone mineral density in grand multiparous women with extended lactation. American journal of obstetrics and gynecology 182: 1371-1377.
- Jørgensen, B. and Vestergaarad, T. 1990. Genetics of Leg Weakness in Boars at Danish Pig Breeding Stations. Acta Agric. Scand. 40: 59-69.
- Keaveny, T.M., Borchers, R.E., Gibson, L.J. and Hayes,
  W.C. 1993. Theoretical analysis of the experimental artifact in trabecular bone compressive modulus.
   Journal of Biomechanics 26: 599-607.
- Kemme, P.A., Jongbloed, A.W., Mroz, Z. and Beynen, A.C. 1997. The efficacy of Aspergillus niger phytase in rendering phytate phosphorus available for absorption in pigs is influenced by pig physiological status. J. Anim Sci. 75: 2129-2138.
- Kirk, R.K., Svensmark, B., Ellegaard, L.P. and Jensen, H.E. 2005. Locomotive Disorders Associated with Sow Mortality in Danish Pig Herds. Journal of Veterinary Medicine 52: 423-428.
- **Lawlor, P. and Lynch, P. 2007.** A review of factors influencing litter size in Irish sows. Irish Veterinary Journal 60: 359-66.
- Mahan, D.C. and Fetter, A.W. 1982. Dietary Calcium and Phosphorus Levels for Reproducing Sows. J. Anim Sci. 54: 285-291.
- Main, D., Clegg, J., Spatz, A. and Green, L. 2000. Repeatability of a lameness scoring system for finishing pigs Veterinary Record 147: 574-576.
- Marchant, J.N. and Broom, D.M. 1996. Effects of dry sow housing conditions on muscle weight and bone strength. ANIMAL SCIENCE-GLASGOW 62: 105-114.

- Maxson, P.F. and Mahan, D.C. 1986. Dietary Calcium and Phosphorus for Lactating Swine at High and Average Production Levels. J. Anim Sci. 63: 1163-1172.
- Nakano, T., Aherne, F., Brennan , J. and Thompson , J.R. 1984. Osteochondrosis in growing swine. Canadian Journal of Animal Science 64: 139-146.
- Nakano, T., Brennan, J.J. and Aherne, F.X. 1987. Leg weakness and osteochondrosis in swine: a review. Canadian Journal of Animal Science 67: 883-901.
- Nimmo, R., Peo, E.J., Moser, B. and Lewis, A. 1981. Effect of level of dietary calcium-phosphorus during growth and gestation on performance, blood and bone parameters of swine. J Anim Sci 52: 1330-42.
- Phillips, A.J., Ostlere, S.J. and Smith, R. 2000. Pregnancy-Associated Osteoporosis: Does the Skeleton Recover? Osteoporosis International 11: 449-454.
- Schenck, E.L., McMunn, K.A., Rosenstein, D.S., Stroshine, R.L., Nielsen, B.D., Richert, B.T., Marchant-Forde, J.N. and Lay, D.C., Jr. 2008. Exercising stall-housed gestating gilts: Effects on lameness, the musculo-skeletal system, production and behavior. J. Anim Sci. jas.2008-1046.
- Shaw, D.T., Rozeboom, D.W., Hill, G.M., Orth, M.W., Rosenstein, D.S. and Link, J.E. 2006. Impact of supplement withdrawal and wheat middling inclusion on bone metabolism, bone strength, and the incidence of bone fractures occurring at slaughter in pigs. J. Anim Sci. 84: 1138-1146.
- **Spencer, G.R. 1979.** Pregnancy and lactational osteoporosis. Animal model: porcine lactational osteoporosis. Am J Pathol. 95: 277-280.
- Tanck, E., Homming, J., Van Lenthe, G. and Huiskies,
  R. 2001. Increase in bone volume fraction precedes architectural adaptation in growing bone. Bone 28: 650-654.
- **Tuyttens, F.A. 2005.** The importance of straw for pig and cattle welfare: A review. Applied Animal Behaviour Science 92: 261-282.
- Uhlorn, H., Dalin, G. and Ekman, S. 1995. Osteochondrosis in wild boar-Swedish Yorkshire crossbred pigs (F2 generation). Acta Vet Scand 36: 41-52.
- Vajda, E.G., Bowman, B.M. and Miller, S.C. 2001. Cancellous and Cortical Bone Mechanical Properties and Tissue Dynamics during Pregnancy, Lactation, and Postlactation in the Rat. Biol Reprod 65: 689-695.